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through the stem by the procedure described previously [1]. After 2 days of incubation in the greenhouse, the young shoot was harvested and analysed.

Extraction and isolation of a new limonoid. Limonoids were extracted from 15 seedlings (82 g) by procedures previously described [1]. The total extract was placed on the top of a silica gel column $(1.6 \times 18 \text{ cm})$. The column was eluted, stepwise, by increasing concns of EtOAc in hexane; 500 ml each of 35, 40, 45, 50 and 55% EtOAc. Fractions containing the compound of interest were still contaminated with nomilin. Therefore, the combined fractions were further purified by a preparative TLC plate, which was developed with solvent system b (see below).

Analysis of limonoids. The isolate obtained above was analysed by TLC, and ¹H NMR and ¹³C NMR spectra. NMR spectra were obtained with a JEOLFT spectrometer, JNM-GX 270 WB. Labelled metabolites obtained from [¹⁴C]acetate feeding were analysed with a Berthold Automatic TLC-linear Analyzer LB 2832. TLC plates were developed with solvent systems: (a) EtOAc-cyclohexane (3:2); (b) CH₂Cl₂-MeOH (97:3) and

(c) EtOAc-CH₂Cl₂ (2:3). Limonoids were revealed by spraying plates with Ehrlich's reagent followed by HCl gas exposure [7].

Acknowledgement—The authors thank Dr. V. P. Maier for his help in the preparation of this manuscript.

REFERENCES

- Hasegawa, S., Bennett, R. D. and Maier, V. P. (1984) Phytochemistry 23, 1601.
- 2. Hasegawa, S. and Herman, Z. (1985) Phytochemistry 24, 1973.
- 3. Herman, Z. and Hasegawa, S. (1981) Phytochemistry 24, 2911.
- Hasegawa, S., Herman, Z. and Ou, P. (1985) Phytochemistry 25, 542.
- 5. Bennett, R. D. and Hasegawa, S. (1981) Tetrahedron 37, 17.
- Hasegawa, S., Bennett, R. D. and Verdon, C. P. (1980) J. Agric. Food Chem. 28, 922.
- 7. Dreyer, D. L. (1965) J. Org. Chem. 30, 749.

Phytochemistry, Vol. 25, No. 8, pp. 1985-1987, 1986. Printed in Great Britain.

0031-9422/86 \$3.00 + 0.00 Pergamon Journals Ltd.

A BIANTHRAQUINONE AND A TRITERPENOID FROM THE SEEDS OF CASSIA HIRSUTA

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Key Word Index—Cassia hirsuta; Leguminosae; seeds; 4.4'-bis(1.3.8-trihydroxy-2-methyl-6-methoxy anthraquinone); pentacyclic triterpernoid sapogenol; isohopane series; $3\beta.16\beta.22$ -trihydroxyisohopane.

Abstract—From the seeds of Cassia hirsuta a new bianthraquinone, 4,4'-bis(1,3,8-trihydroxy-2-methyl-6-methoxy anthraquinone) and a triterpenoid 3β ,16 β ,22-trihydroxyisohopane have been isolated.

INTRODUCTION

Species of Cassia are rich sources of flavonoids [1], anthraquinones [2] and polysaccharides [3] and the plant possesses important medicinal properties [4].

RESULTS AND DISCUSSION

A yellow crystalline pigment (1), $C_{32}H_{22}O_{12}$, ([M]⁺ at m/z 598), mp > 300° (dec.) and a triterpenoid (2), $C_{30}H_{52}O_3$, ([M - H_2O]⁺ at m/z 442), mp 211° were isolated from the ethyl acetate extract of the defatted seeds of Cassia hirsuta.

Compound 1 contained two methyl groups, two methoxyl groups and gave a pink colour with methanolic magnesium acetate [5] for the presence of a chelated (a-hydroxy) hydroxyl group. The solubility of the compound

in 5% aq. sodium bicarbonate and a peak at 3440 cm⁻¹ in its IR spectrum showed the presence of at least one β -hydroxyl group. A medium band at 2910 and 2890 cm⁻¹ indicated the presence of a methoxyl and a methyl group, respectively. Sharp and strong bands at 1630 and 1668 cm⁻¹ suggested the presence of chelated and non-chelated carbonyl groups. The $\lambda_{\text{max}}^{\text{EIOH}}$ at 260, 390 and 440 nm indicated that 1 was a polyhydroxy anthraquinone [6, 7].

On acetylation, a hexaacetyl derivative, C₄₄H₃₄O₁₈ (1a), mp 213°, was obtained. The IR spectrum of 1a showed a phenolic acetate band at 1770 cm⁻¹ and a non-chelated carbonyl at 1668 cm⁻¹. Thus, out of twelve oxygen atoms in compound 1, four occur as quinone carbonyl groups, which are both chelated (1630 cm⁻¹) and non-chelated (1668 cm⁻¹) as revealed from a com-

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parison of the IR spectra of 1 and 1a. This confirmed the presence of a bianthraquinonoid chromophore in 1. The mass spectrum in addition to the molecular ion peak at m/z 598 had strong peaks at [M-17], [M-18], [M-75] and [M-103] exhibiting the loss of water and multiple loss of carbon monoxide typical of hydroxyl anthraquinone [8]. The high molecular weight of 1 ($[M]^+$ at m/z 598) also supported the bianthraquinonoid nature of 1.

Reductive cleavage of 1 with lithium aluminium hydride or with alkaline sodium dithionite [9] gave 1b, $C_{16}H_{12}O_6$, mp 250°, as the only product. Compound 1b was identified as 1,3,8-trihydroxy-2-methyl-6-methoxy anthraquinone by colour reactions [10, 11], UV, IR, NMR, MS and co-chromatography with a natural sample [12]. This suggests that 1 is the dehydro dimer of 1b, $C_{16}H_{12}O_6$.

The ¹H NMR spectrum of 1a showed signals at δ 2.10 (s, 6H, 2 × C-Me), 3.89 (s, 6H, 2 × OMe), 2.45 (s, 6H, 2 × Ac), 2.40 (s, 6H, 2 × Ac), 2.34 (s, 6H, 2 × Ac), 7.65 (d, 2H, J = 2.5 Hz, H-5, H-5') and 6.84 (d, 2H, J = 2.5 Hz, H-7, H-7'). Comparison with the NMR spectrum of the acetate of 1b [signals at δ 2.10 (s, 3H, C-Me), 3.89 (s, 3H, OMe), 7.89 (s, 1H, H-4), 7.67 (d, 1H, J

= 2.5 Hz, H-5), 6.85 (d, 1H, J = 2.5 Hz, H-7), 3.89 (s, 3H), 2.43 (s, 3H), 2.39 (s, 3H) (each due to Ac)] confirms that 1 is a symmetrical dehydro dimer of 1b.

In the ¹H NMR spectrum of 1a a singlet at δ7.89 (assigned to H-4) is absent, showing a C-4, C-4' inter anthraquinone linkage. Thus the compound is 4,4'-bis(1,3,8-trihydroxy-2-methyl-6-methoxy anthraquinone).

Compound 2, $C_{30}H_{52}O_3$ ([M - H_2O]⁺ at m/z 442), mp 211°, gave all the colour tests for a triterpenoid and its IR spectrum showed a peak at 3240 cm⁻¹ for hydroxyl groups. Acetylation gave a monohydroxy diacetate (2a), $C_{34}H_{56}O_5$, ([M]⁺ at m/z 544), mp 225°. The IR spectrum of 2a showed peaks at 3600 (tertiary-OH), 1735, 1755 and 1240-1250 cm⁻¹ (acetoxy). The ¹H NMR spectrum (300 MHz, CDCl₃) of 2a showed signals at δ 0.85 (3H, s, 18α -Me); 1.05 (3H, s, 4β -Me); 1.1 (3H, s, 4α -Me); 1.16 (3H, s, 10β -Me) and 1.19 (6H, s, 8β - and 14α -Me) for six tertiary methyl groups [13, 14]. Two methyl groups in a hydroxyisopropyl side chain (Me-C-(OH)-Me) appeared at δ 1.50 (3H, s, C-29) and 1.52 (3H, s, C-30). The signals at δ 2.06 (3H, s) and 2.08 (3H, s) were due to the two acetoxyl methyl groups (2 × OAc). The singlet at δ 2.92 (1H) which disappeared with D₂O was due to the tertiary hydroxyl

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groups. The 3-axial (α) proton appeared as a doublet at δ 4.2 (1H, J=7.5 Hz). A weak multiplet at δ 5.55 (1H) for the 16β (axial) proton, as is usually observed in 3β - and 16β -acetoxy triterpenes [15].

The mass spectrum of 2a showed a molecular ion peak at m/z 544 together with peaks at m/z 526 [M - H₂O]⁺, 466 [M - H₂O - HOAc]⁺, 426 [M - HOAc - 58]⁺, 424

[M-2 × HOAc]⁺, 249, 59.

The oxidation of 2 with CrO_3 -pyridine [16] gave a monohydroxy diketone (2b), mp 232°, and a dihydroxy monoketone (2c), $C_{30}H_{50}O_3$, mp 228°. The diketone 2b readily isomerized to the α , β -unsaturated ketone 2d by refluxing with EtOH-HCl (5%); $C_{30}H_{46}O_2$, mp 234°. The easy isomerization indicated the presence of two other hydroxyl groups at C-16 (secondary) and C-22 (tertiary), respectively. Easy acetylation indicated the two secondary hydroxyl groups of 2 appeared to be equitorial (β). On considering all the above facts compound 2 may be represented at 3β , 16β , 22-trihydroisohopane. This is also confirmed by mp, mmp and co-chromatography with an authentic sample obtained from the flowers of *Mollugo hirta* [17].

EXPERIMENTAL

Dried and crushed seeds were extracted with refluxing petrol and then with refluxing EtOAc and concd. The concd extract contained five spots (silica gel TLC) which were separated by a silica gel column using different solvents. The EtOAc fraction gave a single spot on TLC (silica gel, C_0H_6 -EtOAc, 4:1), yellow compound (1) crystallized from EtOAc-petrol (1:3) mp > 300°. (Calc.: C, 64.21; H, 3.68. Found: C, 64.20; H, 3.70%). UV λ_{\max}^{EtOH} nm: 240, 290, 430. IR ν_{\max}^{KBr} cm⁻¹: 3340, 2960, 2800, 1675, 1630, 1150. The acetate was prepared with Ac₂O-pyridine mp 213°. UV λ_{\max}^{EtOH} nm: 260, 290, 440. IR ν_{\max}^{KBr} cm⁻¹: 1770, 1663, 1668, 1452, 1348.

Vacuum dried product 1 (100 mg) was dissolved in aq. 1 N $\rm Na_2CO_3$ (200 ml) and a soln of sodium dithionite (20 mg) in $\rm H_2O$ (100 ml) was added through a filter paper. After gentle shaking for 5 min a vol. of 1 N HC1 equivalent to the $\rm Na_2CO_3$ present was cautiously added. The yellow solid (1b) was precipitated and crystallized from HOAc, mp 250°.

The C₆H₆-MeOH (4:1) eluate of the silica gel column gave a single spot on TLC (silica gel, EtOAc) of compound 2, crystallized with CHCl₃-MeOH, mp 211°. (Calc.: C, 78.21; H, 11.38. Found: C, 78.01%; H, 11.16%.) The acetate was prepared with

Ac₂O-pyridine, mp 225°. IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3600, 1755, 1735, 1240, 1250. ¹H NMR (300 MHz, CDCl₃): δ 0.85 (3H, s, Me), 1.05 (3H, s, Me), 1.10 (3H, s, Me), 1.16 (3H, s, Me), 1.19 (6H, s, 2 × Me), 1.50 (3H, s, H-29), 1.52 (3H, s, H-30), 2.06 (3H, s) and 2.08 (3H, s), both due to acetoxy methyl groups (2 × OAc), 4.2 (1H, d, J = 7.5 Hz), 5.55 (s, 1H).

Oxidation of compound 2. A cold soln of 2 in dry pyridine (5 ml) was added to a slurry of CrO₃-pyridine complex at 0° with stirring and left overnight at room tmep. The reaction mixture was poured into ice and the recovered product crystallized from petrol-CHCl₃, mp 232°.

α,β-Unsaturated ketone 2d from 2b. Compound 2b was dissolved in dry EtOH (4 ml) and dry EtOH-HCl (5%, 4 ml) was added. The soln was refluxed for 3 hr. The reaction mixture was poured into ice and the recovered product crystallized from MeOH, mp 234°.

Acknowledgements—One of us (Janhavi Singh) is grateful to C.S.I.R., India for the award of J.R.F. Dr. P. D. Satsangi, The Pennsylvania State University, U.S.A., is thanked for mass and NMR spectra.

REFERENCES

- 1. Tiwari, R. D. and Singh, J. (1978) Planta Med. 34, 319.
- 2. Tiwari, R. D. and Singh, J. (1978) Phytochemistry 18, 347.
- 3. Gupta, P. C. and Dubey, P. (1978) Carbohydr. Res. 72, 151.
- Chopra, R. N., Chopra, I. C., Hande, K. L. and Kapur, L. D. (1958) Indigenous Drugs of India, Vol. 98, p. 499. Dhar, U.N.
- 5. Shibata, S. J. (1941) Pharm. Soc. Japan 61, 320.
- 6. Bowie, J. H. (1962) Aust. J. Chem. 15, 336.
- 7. Birch, A. J. and Massy, R. A. (1957) J. Chem. Soc. 2215.
- Bowie, H. H., Hoffman, P. J. and White, P. Y. (1970) Tetrahedron 26, 1163.
- 9. Howard, B. H. and Raistrick, H. (1954) Biochem. J. 56.
- 10. Somogyi, J. (1952) Biochem. J. 19, 195.
- Shibata, S., Takido, M. and Tanaka, O. (1950) J. Am. Chem. Soc. 72, 2789.
- 12. Tiwari, R. D. and Singh, J. (1983) Z. Naturforsch 38h, 1136.
- 13. Lehn, J. M. and Vystrcil, A. (1963) Tetrahedron 19, 1733.
- 14. Corbett, R. E. and Smith, R. A. J. (1967) J. Chem. Soc. C 1622.
- Shamma, M., Glick, R. E. and Mumma, R. O. (1962) J. Org. Chem. 24, 4512.
- Poos, G. I., Arth, G. E., Beyler, R. E. and Sarett, L. H. (1953) J. Am. Chem. Soc. 75, 422.
- Choudhury, M. K. and Chakarabarti, P. (1979) Phytochemistry 18, 1363.